

## Growth kinetics of *Pseudomonas fluorescens* in different media and evaluation of shelf life

ANDREA SUSAN BABY<sup>1</sup>\*, GURUDATT M. HEGDE<sup>1</sup>, SHAMARAO JAHAGIRDAR<sup>1</sup> AND C. R. PATIL<sup>2</sup>

<sup>1</sup>Department of Plant Pathology, <sup>2</sup>Department of Microbiology, College of Agriculture, Dharwad

University of Agricultural Sciences, Dharwad - 580 005, India

\*E-mail: andreasusanbaby@gmail.com

(Received: November, 2023 ; Accepted: July, 2025)

DOI: 10.61475/JFS.2025.v38i3.16

**Abstract:** Biopesticides open up a wide scope for restoration of environment that has been damaged through continuous use of chemicals. The study focused on assessing the growth kinetics of *Pseudomonas fluorescens* in various liquid media and evaluating their shelf life. Seven media were analysed for their ability to assist the growth and multiplication of the bacterium. Among the various media evaluated, Trypticase Soya broth (TSB) was considered as the effective basal medium owing to its ability to maintain high population, lesser generation time and lesser cost. Further, it was formulated with various amendments like glycerol and PVP (cell protectants), CMC (adjuvant) and Tween 20 (surfactant) and the formulation (TSB+ Glycerol 2%+ PVP 1%+ CMC 0.05%+ Tween-20 0.5%) was found to be promising in sustaining the higher population of *Pseudomonas fluorescens* up to 240 days of storage.

**Key words:** Biopesticide, Cell amendments, Liquid formulation, *Pseudomonas fluorescens*, Shelf life

### Introduction

Integration of plant growth promoting bacteria in sustainable agriculture has been practiced over several decades. Even though there are ample of promising research regarding the antagonism of bioagents under lab conditions, the field efficacy offered by them is yet a puzzle to be solved. This is often attributed to the various environmental stresses they have to cope up with, the major ones being adaptation to introduced niche, competition of already established organisms and inability to multiply. This emphasizes the need of formulating the bioagents for better performance in field conditions (Bashan *et al.*, 2014).

Majority of microbial formulations in India at present are mostly carrier based, and are widely used in that form. But these carrier-based formulations have many shortcomings, the prominent among them being the reduced shelf life of the produce, higher contamination, intolerance towards UV rays and high temperature, bulkiness, blockage of nozzles in drip irrigation and lower field performance. In this context, a better alternative is the liquid formulations that contain the cultures or cell suspensions of the microbial organisms, amended with surfactants, adjuvants, stabilizers, preservatives, etc., that can sustain the viability and support its growth by providing necessary nutrients (Vora *et al.*, 2008; Teixidó *et al.*, 2022). Apart from showing high cell count, they also exhibited increased shelf life of up to one year, zero contamination, improved protection against biotic and abiotic stresses which boosted the field efficacy compared to peat-based inoculants (Singleton *et al.*, 2002).

Therefore, the aim of this research was to develop a formulation of *Pseudomonas fluorescens*, which is regarded as an efficient antagonist in curbing the pathogens (Panpatte *et al.*, 2016; Mehmood *et al.*, 2023), that can maintain the shelf life and potency.

### Material and methods

*Pseudomonas fluorescens* (IOF) strain (NAIMCC-B-01981) from the Institute of Organic Farming, University of Agricultural Sciences, Dharwad, was used to develop the formulation.

#### Selection of suitable growth medium by developing the growth curve

Seven different media *viz.*, King's B broth (KB), Nutrient Broth (NB), Trypticase Soya Broth (TSB), Luria- Bertani Broth (LB), *Pseudomonas* (Fluorescein) broth (PF), Minimal salt medium (M9) and Potato Dextrose broth (PDB), were taken to evaluate the growth of *P. fluorescens*. One day old culture of *Pseudomonas fluorescens* was inoculated @ ten per cent to different media and were incubated at room temperature (100 rpm). Growth curve of *P. fluorescens* was developed by enumerating the number of viable cells by standard plate count technique on alternate hours for 24 h in different media. The petri plates were incubated at  $30 \pm 1^\circ\text{C}$  for 48 h and the number of colonies were calculated using the formula: average no. of colonies  $\times$  dilution factor/ Volume taken for dilution. Growth curve was obtained by plotting colony forming units as a function of time. The data were used to calculate the generation time of *Pseudomonas fluorescens* in different culture media, using the formula (Madigan *et al.*, 2015):

$$\text{Generation time (g)} = t/n$$

Where,

t- time interval in hour, between-two estimates

n-Number of generations ( $n = 3.3 (\log N - \log N_0)$ )

(N = the final cell count;  $N_0$  is initial cell count)

The best basal medium for the formulation was selected based on criteria like minimum generation time, the highest population achieved and the time taken for the same, contamination level and cost economics.

Table 1. Population of *Pseudomonas fluorescens* in different media

Media	Population in log transformed values ( $10^0$ cfu/ml)												
	0 <sup>th</sup> h	2 <sup>nd</sup> h	4 <sup>th</sup> h	6 <sup>th</sup> h	8 <sup>th</sup> h	10 <sup>th</sup> h	12 <sup>th</sup> h	14 <sup>th</sup> h	16 <sup>th</sup> h	18 <sup>th</sup> h	20 <sup>th</sup> h	22 <sup>nd</sup> h	24 <sup>th</sup> h
KB	8.078 (0.012)	8.079 (0.012)	8.174 (0.015)	8.195 (0.016)	8.700 (0.050)	9.050 (0.112)	9.213 (0.163)	9.277 (0.189)	9.414 (0.260)	9.588 (0.387)	9.845 (0.700)	9.715 (0.519)	9.560 (0.363)
NB	8.037 (0.011)	8.045 (0.011)	8.146 (0.014)	8.177 (0.015)	8.68 (0.048)	9.018 (0.104)	9.150 (0.141)	9.235 (0.172)	9.342 (0.220)	9.495 (0.313)	9.713 (0.516)	9.486 (0.306)	9.332 (0.215)
TSB	8.082 (0.012)	8.090 (0.012)	8.185 (0.015)	8.198 (0.016)	8.710 (0.051)	9.152 (0.142)	9.499 (0.316)	9.595 (0.393)	9.699 (0.500)	9.889 (0.770)	10.175 (1.496)	10.209 (1.618)	10.300 (1.990)
PF	7.940 (0.009)	8.017 (0.010)	8.028 (0.011)	8.056 (0.015)	8.293 (0.019)	8.491 (0.031)	8.572 (0.037)	8.653 (0.045)	8.776 (0.059)	8.890 (0.078)	9.157 (0.143)	9.390 (0.245)	9.588 (0.387)
LB	8.024 (0.011)	8.053 (0.011)	8.093 (0.012)	8.154 (0.014)	8.572 (0.037)	8.999 (0.099)	9.123 (0.132)	9.225 (0.168)	9.374 (0.236)	9.500 (0.316)	9.623 (0.420)	9.704 (0.506)	9.743 (0.553)
PDB	7.603 (0.004)	7.713 (0.005)	7.722 (0.006)	7.800 (0.011)	8.050 (0.018)	8.271 (0.021)	8.322 (0.026)	8.411 (0.040)	8.596 (0.041)	8.609 (0.064)	8.810 (0.108)	9.033 (0.111)	9.047
M9	7.271 (0.002)	7.301 (0.002)	7.325 (0.003)	7.470 (0.005)	7.679 (0.011)	8.033 (0.020)	8.206 (0.022)	8.340 (0.032)	8.512 (0.040)	8.599 (0.050)	8.698 (0.068)	8.835 (0.077)	8.885

Note: KB- King's B Broth; NB- Nutrient Broth; TSB-Trypticase Soya Broth; PF- *Pseudomonas* (Fluorescein) broth; LB- Luria Bertani Broth; PDB- Potato Dextrose broth; M9- Minimal salt medium \*Values in parenthesis represent original CFU (Colony forming units $\times 10^{10}$ )

### Preparation of liquid formulations of *Pseudomonas fluorescens* and assessment of shelf life

Once the best basal medium was selected, it was amended with two concentrations each of cell protectants (Poly vinyl pyrrolidone (PVP) (@ 1% and 2%) and glycerol (@ 2% and 1%), adjuvant (carboxy methyl cellulose (CMC) (@ 0.05 % and 0.10%) and surfactant (Tween-20 (@ 0.5% and 1.0%) (Santhosh, 2015; Biradar and Santhosh, 2018; Gopi *et al.*, 2019). Thus, a total of 16 formulations with all the combinations of these amendments were evaluated and shelf life was determined by serial dilution and plate count method for following eight months.

### Results and discussion

Among the seven media evaluated, the population observed in Trypticase Soy broth ( $1.99 \times 10^{10}$  CFU ml $^{-1}$ ) was found to be highest at the end of 24<sup>th</sup> h of observation, followed by the population in Luria Bertani broth ( $0.553 \times 10^{10}$  CFU ml $^{-1}$ ). In KB and NB broths, the population increased till 20<sup>th</sup> h in incubation, but there was a sudden decline in population afterwards. The least population at 24<sup>th</sup> h ( $0.077 \times 10^{10}$  CFU ml $^{-1}$ ) was recorded with medium M9 (Table 1).

Trypticase Soya broth (TSB) exhibited the least generation time of 1.26 h, followed by King's B broth (1.41 h). It was comparable with that of Luria-Bertani Broth (1.42 h) and Nutrient Broth (1.43 h). The highest generation time for this bacterium was recorded in *Pseudomonas* (Fluorescein) broth (2.77 h). When the cost of the media was taken into consideration, *Pseudomonas* (Fluorescein) broth was the costliest, recording 15,706 ₹ /100 l, followed by King's B broth (12,714 ₹ /100 l). Minimal salt medium (M9) was the least costly. (2,201.78 ₹ /100 l) (Table 2). Thus, Trypticase Soya broth was finally selected as the basal medium for shelf life studies, owing to its least generation time, lesser cost and ability to maintain higher population. The growth curves of the bacterium in different media are depicted in Fig 1. Vyas *et al.* (2014), revealed that the highest population of viable colonies of phosphate solubilizing bacteria, *Pseudomonas trivialis* BIHB 745 were maintained in Trypticase soy Broth, among the seven different media (King's B medium, Trypticase soya broth, TSB with 2% methanol, Nutrient broth, glucose yeast extract medium, *Pseudomonas* medium A and *Pseudomonas* medium 2) evaluated. The use of TSB for mass production of *P. fluorescens* was further authenticated by Rajeshwari and Appanna (2021), who found it ideal for maintaining the optimal growth of the bacteria.

Table 2. Properties of various media used in the study

Media	Generation time (h)	Cost of media required to prepare 100 l (₹)	Generation time, merits and demerits of the media
King's B (KB)	1.41	12,714	Generation time is short, but sudden decline in population, high cost of medium
Nutrient Broth (NB)	1.43	9175	Short generation time, but sudden decline in population
Trypticase Soya Broth (TSB)	1.26	9796	Maintained high population, shortest generation time, reasonable cost of medium
<i>Pseudomonas</i> (Fluorescein) the broth (PF)	2.77	15,706	High cost of the medium and highest generation time among all media
Luria- Bertani Broth (LBB)	1.42	11,495	Short generation time, but lesser population and costlier as compared to
TSB			It's a general medium used for the growth of fungi rather than bacteria, had higher generation time and lower population
Potato Dextrose broth (PDB)	2.56	2608	Lesser population, and more generation time
Minimal salt medium (M9)	2.14	2201.78	

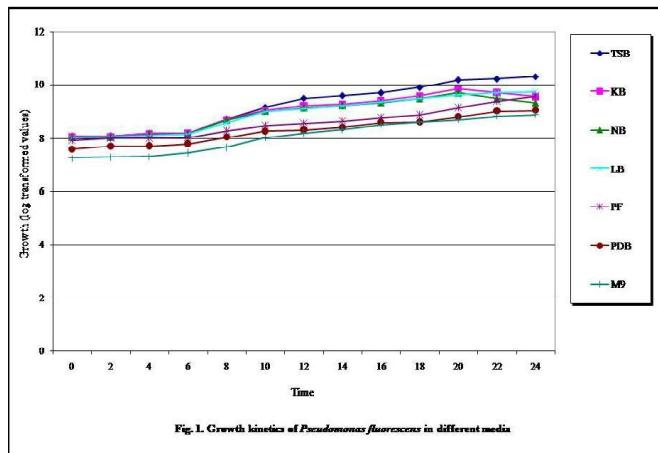


Fig 1. Growth kinetics of *Pseudomonas fluorescens* in different media

### Preparation of liquid formulations of *Pseudomonas fluorescens* and assessment of shelf life

Various amendments were incorporated to the basal medium (TSB) at different concentrations and the population (CFU ml<sup>-1</sup>) were estimated at monthly intervals (Table 3). The results revealed that, the viable count of colonies increased from 0<sup>th</sup> day and reached a maximum by 30<sup>th</sup> day of storage. The highest population observed (on 30<sup>th</sup> day) was  $48.33 \times 10^{10}$  CFU ml<sup>-1</sup>, in formulation 9. Later, the population declined gradually over the months of storage. Even at the end of eight months, the

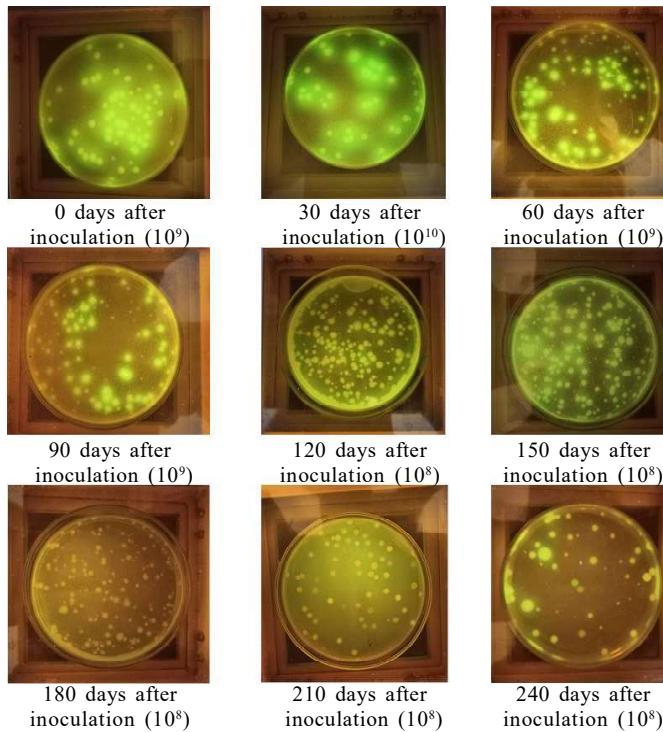
highest population was maintained in formulation 9 (TSB+ Glycerol 2%+ PVP1%+ CMC0.05%+ Tween-20 0.5%) ( $66 \times 10^8$  CFU ml<sup>-1</sup>), followed by formulation 1 (TSB+ Glycerol 1%+ PVP 1%+ CMC 0.05%+ Tween-20 0.5%) ( $52 \times 10^8$  CFU ml<sup>-1</sup>). The lowest population among the formulations at 240<sup>th</sup> day, was found in formulation 5 (TSB+ Glycerol 1%+ PVP 2%+ CMC 0.05%+ Tween-20 0.5%) ( $3 \times 10^8$  CFU ml<sup>-1</sup>). The population of bacteria in unamended (control) TSB ( $28 \times 10^4$  CFU ml<sup>-1</sup>) was several folds lower as compared to the amended formulations. Among the checks, liquid based market sample of the antagonist maintained the population till 240 days ( $4 \times 10^8$  CFU ml<sup>-1</sup>), while talc-based checks could not maintain requisite population beyond 180 days. The viable colonies obtained on different days of storage for formulation 9 is represented in Fig 2.

The amendments added to the medium had a positive effect in protecting the cells and prolonging the shelf-life when compared to the unamended broth and talc formulations. The cell protectants *viz.*, glycerol and PVP retained the water content, maintained the viability (Manikandan *et al.*, 2010; Biradar and Santhosh, 2018). The adjuvant CMC retained the integrity of cells by maintaining a thick consistency of the medium, while Tween-20 reduced the surface tension of the medium, as a surfactant (Daniel *et al.*, 2013; Biradar and Santhosh, 2018). Manikandan *et al.* (2010) have documented the population count of *P. fluorescens* as  $9.5 \times 10^7$  CFU ml<sup>-1</sup>, in Nutrient broth amended with 10 mM glycerol. Similarly, shelf life was maintained till 210<sup>th</sup> day of storage when nutrient broth

Table 3. Shelf life of *Pseudomonas fluorescens* at different days of storage in trypticase soya broth

Formulation No.	Composition of formulations	Colony forming units (CFU $\times 10^{10}$ ) per ml of formulation at different DAI								
		0	30	60	90	120	150	180	210	240
1	TSB+ G(1%)+P(1%)+C(0.05%)+T(0.5%)	7.23	47.00	8.60	8.07	1.75	1.63	1.26	0.89	0.52
2	TSB+ G(1%)+P(1%)+C(0.05%)+T(1.0%)	6.77	40.67	7.43	6.24	1.51	1.33	0.90	0.38	0.11
3	TSB+ G(1%)+P(1%)+C(0.1%)+T(0.5%)	7.20	45.00	8.83	7.65	1.72	1.51	1.17	0.66	0.49
4	TSB+ G(1%)+P(1%)+C(0.1%)+T(1%)	6.63	36.00	6.90	5.43	1.28	1.15	0.70	0.10	0.07
5	TSB+ G(1%)+P(2%)+C(0.05%)+T(0.5%)	6.17	27.33	5.07	2.40	0.51	0.34	0.26	0.05	0.03
6	TSB+ G(1%)+P(2%)+C(0.05%)+T(1%)	6.13	29.33	5.27	2.73	0.61	0.47	0.29	0.05	0.04
7	TSB+ G(1%)+P(2%)+C(0.1%)+T(0.0%)	7.07	42.67	7.93	7.06	1.63	1.37	0.99	0.46	0.26
8	TSB+ G(1%)+P(2%)+C(0.1%)+T(1.0%)	6.20	34.67	6.10	3.20	1.03	0.79	0.56	0.09	0.06
9	TSB+ G(2%)+P(1%)+C(0.05%)+T(0.5%)	7.43	48.33	9.57	8.50	1.97	1.84	1.36	1.05	0.66
10	TSB+ G(2%)+P(1%)+C(0.05%)+T(1%)	7.13	44.00	8.27	7.68	1.68	1.47	1.12	0.60	0.34
11	TSB+ G(2%)+P(1%)+C(0.1%)+T(0.5%)	6.87	42.33	7.57	6.54	1.57	1.42	1.04	0.53	0.29
12	TSB+ G(2%)+P(1%)+C(0.1%)+T(1%)	6.30	31.67	5.73	3.07	0.85	0.56	0.32	0.06	0.05
13	TSB+ G(2%)+P(2%)+C(0.05%)+T(0.5%)	6.73	39.33	7.32	6.10	1.47	1.29	0.85	0.32	0.09
14	TSB+ G(2%)+P(2%)+C(0.05%)+T(1%)	6.33	35.33	6.47	3.43	1.15	0.99	0.60	0.09	0.06
15	TSB+ G(2%)+P(2%)+C(0.1%)+T(0.5%)	6.50	37.33	7.19	5.87	1.36	1.25	0.76	0.25	0.08
16	TSB+ G(2%)+P(2%)+C(0.1%)+T(1%)	6.27	31.33	5.40	2.80	0.75	0.65	0.41	0.08	0.05
Unamended TSB broth		6.13	2.50	0.80	0.54	0.13	0.09	0.04	0.02	0.00
<i>P. fluorescens</i> - Talc based formulation (IOF strain) (check)		0.34	0.44	0.38	0.33	0.12	0.09	0.04	0.02	0.00
<i>P. fluorescens</i> - Talc based formulation (market sample) (check)		0.28	0.36	0.33	0.26	0.09	0.06	0.03	0.00	0.00
<i>P. fluorescens</i> - Liquid based formulation (market sample) (check)		0.85	0.88	0.60	0.56	0.15	0.12	0.10	0.04	0.04
S.Em $\pm$		0.07	0.06	0.10	0.07	0.01	0.01	0.01	0.01	0.01
C.D @ 1%		0.28	0.22	0.37	0.26	0.04	0.04	0.04	0.03	0.03
C.V. (%)		2.22	2.58	2.87	2.67	1.75	1.80	2.43	3.72	5.07

Note: TSB- Trypticase Soya Broth; G- Glycerol; P- Poly vinyl pyrrolidone; C- Carboxy methyl cellulose; T-Tween-20  
DAI-Days after inoculation



## References

Bashan Y, de-Bashan L E, Prabhu S R and Hernandez J, 2014, Advances in plant growth-promoting bacterial inoculant technology: formulations and practical perspectives (1998-2013). *Plant Soil*, 378: 1-33.

Biradar P B J and Santhosh G P, 2018, Cell protectants, adjuvants, surfactant and preservative and their role in increasing the shelf life of liquid inoculant formulations of *Pseudomonas fluorescens*. *International Journal of Pure and Applied Bioscience*, 6(4): 116-122.

Daniel L A E, Venkateshwarlu B, Suseelandra D, Pavankumar G, Mirhassan A S K, Meenakshi T, Uzma S, Sravani P and Lakshmi N M, 2013, Effect of polymeric additives, adjuvants, surfactants on survival, stability and plant growth promoting ability of liquid bioinoculants. *Journal of Plant Physiology and Pathology*, 1(2): 1-5.

Dobhal A and Hegde G M, 2021, Shelf life study of *Pseudomonas fluorescens* in liquid formulations. *Journal of Farm Sciences*, 34(1): 84-86.

Gopi G K, Meenakumari K S, Nysanth N S, Subha P, 2019, An optimized standard liquid carrier formulation for extended shelf-life of plant growth promoting bacteria. *Rhizosphere*, 11: 1-4.

Madigan M T, Martinko J M, Bender K S, Buckley D H and Stahl D A, 2015, Brock biology of microorganisms. Pearson Publishers, San Francisco, USA, pp. 149-156.

Manikandan R, Saravanakumar D, Rajendran L, Raguchander T and Samiyappan R, 2010, Standardization of liquid formulation of *Pseudomonas fluorescens* Pf1 for its efficacy against *Fusarium* wilt of tomato. *Biological Control*, 54(2): 83-89.

Mehmood N, Saeed M, Zafarullah S, Hyder S, Rizvi Z F, Gondal A S, Jamil N, Iqbal R, Ali B, Ercisli S and Kupe M, 2023, Multifaceted impacts of plant-beneficial *Pseudomonas* spp. in managing various plant diseases and crop yield improvement *American Chemical Society Omega*, 8(25): 22296-22315.

Panpatte D G, Jhala Y K, Shelat H N, and Vyas R V, 2016, *Pseudomonas fluorescens*: A Promising biocontrol agent and PGPR for sustainable agriculture. In: *Microbial Inoculants in Sustainable Agricultural Productivity* (Eds. Singh D P, Singh H B and Prabha R), Springer, New Delhi, pp. 257-270.

Paranthaman L and Chellappan G, 2020, Influence of storage period of *P. fluorescens* (TPf12) liquid formulation on *F. oxysporum* f. sp. *lycopersici*. *Journal of Entomology and Zoology Studies*. 8(6): 1379-1386.

Rajeshwari R and Appanna V, 2021, Mass production and formulation of antagonists. In: *Biopesticides in Horticultural Crops* (Eds. Rajeshwari R and Appanna V), CRC Press, Florida, United States, pp. 216-226.

Santhosh G P, 2015, Formulation and shelf life of liquid biofertilizer inoculants using cell protectants. *International Journal of Researches in Biosciences, Agriculture and Technology*, 2: 243-247.

Singleton P, Keyser H, Sande E, 2002, Development and evaluation of liquid inoculants. In: *Inoculants and nitrogen fixation of legumes in Vietnam* (Ed. Herridge D), ACIAR: Canberra, Australia, 109e, pp. 52-66.

Teixidó N, Usall J and Torres R, 2022, Insight into a successful development of biocontrol agents: production, formulation, packaging and shelf life as key aspects. *Horticulturae*, 8(4): 1-16.

Vyas P, Rahi P, Chadha B S and Gulati A, 2014, Statistical optimization of medium components for mass production of plant growth-promoting microbial inoculant *Pseudomonas trivialis* BIHB 745 (MTCC5336). *Indian Journal of Microbiology*, 54(2): 239-241.

Vora M S, Shelat H N and Vyas R V, 2008, Liquid biofertilizers: a new vista. In: *Handbook of biofertilizers and microbial pesticides*, Satish Serial Publishing House, New Delhi, pp. 87-90.

amended with 2% PVP was used (Paranthaman and Chellappan, 2020) and till 12 months when canola oil-based formulation was used to grow *P. fluorescens* (Dobhal and Hegde, 2021). In all the above citations, the population observed at the end of storage period was highly significant over unamended broth or talc-based formulations, emphasizing the fact that, the liquid formulations are more efficient.

## Conclusion

Formulating the bioagents is a crucial step in attaining the full potential of any antagonist. When this aim was kept at the fore, the investigation resulted in formulating a liquid biopesticide of *P. fluorescens* that could maintain the shelf life better than the talc formulations. Trypticase Soya Broth was selected as the basal medium and was amended with various additives to protect the cells from desiccation and to prolong the viability. The study concluded that the developed formulation of *P. fluorescens* (TSB+ Glycerol 2%+ PVP 1%+ CMC 0.05%+ Tween-20 0.5%) was highly efficient in maintaining the shelf life of the antagonist and thus, can be utilized in mass production.